Original Research Article

Surgical site infection in a coastal tertiary care teaching hospital

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	International Archives of Integrated Medicine, Vol. 5, Issue 1, January, 2018. Copy right © 2018, IAIM, All Rights Reserved.		
	Available online at <u>http://iaimjournal.com/</u>		
Jan 1	ISSN: 2394-0026 (P)	ISSN: 2394-0034 (O)	
IAIM	Received on: 26-12-2017	Accepted on: 05-01-2018	
	Source of support: Nil	Conflict of interest: None declared.	
How to cite this article: Mohamed Jan, Haja Abdul Nazeer M.J. Surgical site infection in a coastal			
tertiary care teaching hospital. IAIM, 2018; 5(1): 100-106.			

Abstract

Introduction: Surgical site infections are one among the commonly reported nosocomial infections. The present study was aimed at determining the possible risk factors involved in development of SSIs and the microorganisms responsible for SSIs with their antibiogram in a rural tertiary care teaching hospital.

Materials and methods: This was a prospective observational study conducted in the Department of Surgery in association with the Department of Microbiology at Vinayaka Missions Medical College and Hospital over a period of six months. Detailed history such as age, sex, operation category (elective/ emergency), history of diabetes were recorded. Swabs were collected from clinically suspected of surgical site infections. Specimens collected were subjected to standard microbiological procedures. Antimicrobial susceptibility of the isolates was determined by Kirby Bauer disk diffusion method on Muller Hinton agar plates according to Clinical and Laboratory Standard Institute (CLSI) guidelines.

Results: A total of 342 patients underwent surgery during the study period. 59 cases were clinically diagnosed of surgical site infection, out of which 55 patients' samples yielded growth and accounted for 16% infection rate. The incidence of infection was high in case of emergency surgeries (22%) compared to elective surgeries (13%). SSI rate was low in clean surgeries (7%) whereas contaminated and dirty wounds showed 43% and 75% respectively. Staphylococcus aureus (27.5%) was the predominant pathogen isolated. Linezolide was the most susceptible antibiotic against Gram positive cocci. Second predominant organism was Escherichia coli 14 (20.2%). and showed good susceptibility to imipenem, cefeperazone/ sulbactum and piperacillin/ tazobactum including ESBL producers.

Conclusion: In this study, surgical site infection rate was 16%. Staphylococcus aureus (27%) was the most frequently isolated bacteria followed by E.coli (20%). Good susceptibility observed towards linezolide against Gram positive cocci. Imipenem, cefeperazone/ sulbactum and piperacillin/ tazobactum were effective against Gram negative bacilli.

Key words

Surgical site infection, Stahylococcus aureus, Linezolide.

Introduction

Post-operative wound infection is defined as that infection presenting up to 30 days after a surgical procedure if no prosthetic is placed and up to 1 year if a prosthetic is implanted in the patient [1]. Centers for Disease Control (CDC) revised its definition of 'wound infection', by creating the definition, 'surgical site infection' (SSI) to prevent the confusion between the infection of a surgical incision and the infection of a traumatic wound [2]. Most of the SSIs are superficial, but even so, they contribute greatly to the morbidity and the mortality which are associated with the surgeries [3]. SSI's are the third most frequently reported nosocomial infections, accounting for 14-16% of all the infections. Worldwide estimates of SSI have varied from 0.5-15% [4-6]. Studies in India have consistently shown higher rates up to 38% [7, 8].

Various factors are responsible for developing SSIs. These include the degree of microbial contamination of the operation site indicated by wound class as clean, clean contaminated, contaminated and dirty, and also by patient age, length of surgery, pre-operative shaving of the operative site, hypothermia and comorbidities e.g. diabetes and obesity [9]. The probability of wound infection is determined largely by the interaction of the microbial burden, local wound conditions, and the patient's systemic host defenses. The conditions of antimicrobial therapy, both prophylactically and therapeutically, can only be defined when these other factors are under control [10].

Thus, the identification of factors that cause or predict these infections continues to be an important area of research. A wide variety of aerobic and anaerobic species of bacteria may be present, either singly or in combination [11]. Multi drug resistant bacteria (MDR) is one of the major threats due to widespread use of antimicrobial drugs. These bacteria exhibit resistance to two or more classes of antibiotics. In recent years, multi drug resistant bacteria such as, Methicillin resistant Staphylococcus aureus extended-spectrum-β-lactamases (MRSA), enterobacteria (ESBL) producing and vancomycin resistant enterococci (VRE) have become common particularly among hospitalized Prevalence antibacterial patients. and susceptibility patterns of MDR organisms are important for choosing appropriate empirical therapy especially to treat surgical site infections [12].

The present study was aimed at determining the possible risk factors involved in development of SSIs and the microorganisms responsible for SSIs with their antibiogram in a rural tertiary care teaching hospital.

Materials and methods

This was a prospective observational study conducted in the Department of Surgery in association with the Department of Microbiology at Vinayaka Missions Medical College and Hospital over a period of six months (January 2017 to June 2017) on all patients who underwent surgical procedures during study period. Verbal informed consent was obtained from all the patients. Detailed history such as operation age, sex. category (elective/ emergency), history of diabetes were recorded. The operations were classified, depending on the degree of contamination, as Clean, Clean-Contaminated, and Contaminated employing the

American College of Surgeons' Committee for Control of Surgical Infections guidelines [13].

The cumulative incidence of SSIs was expressed as infection rate - the number of patients with SSIs per 100 operated patients. Patients less than 14 years, Patients undergoing re-operations and those with open wounds for desloughing were excluded. Specimens were collected from the patients in the form of swabs from suspected surgical site infection (8-10 days and then 2-4 weeks after the surgery from the wounds having serous or purulent discharge, showing signs of inflammation). Two swabs were collected from each patient and subjected to standard microbiological procedures. Antimicrobial susceptibility of the isolates was determined by Kirby Bauer disk diffusion method on Muller Hinton agar plates according to Clinical and Laboratory Standard Institute (CLSI) guidelines [14].

Results

A total of 342 patients underwent surgery during the study period. 59 cases were clinically diagnosed of surgical site infection, out of which 55 patients' samples yielded growth and accounted for 16% infection rate. Surgical site infection was predominantly observed among patients above 65 years and accounted for 46.42% (**Table - 1**). The description of variables and its association with SSI is presented in **Table - 2**.

Table – 1: Age and sex	wise distribution	of surgical site infections.

Age in years	Male (n=230)	Female (n=112)	Total (n=342)	SSIs (%)
15-25	32	16	48	3(6.25%)
26-35	38	16	54	3(6.25%)
36-45	35	32	67	7(10.44%)
46-55	57	20	77	12(15.54%)
56-65	48	20	68	17(25%)
>65	20	8	28	13(46.42%)

<u>**Table – 2**</u>: Factors associated with surgical site infections.

Operation category	Number	SSIs (%)	
Emergency	97	22(22.6%)	
Elective	245	33(13.46%)	
Type of operation			
Clean	189	15(7.9%)	
Clean contaminated	105	14(13.3%)	
Contaminated	32	14(43.7%)	
Dirty	16	12(75.2%)	
Co morbidities			
Diabetic	92	22(23.9%)	
Non diabetic	250	33(13.2%)	

Out of 59 specimens collected from patients clinically suspected of SSIs, 55 samples yielded growth. Monomicrobial growth was observed in 45 specimens and 10 specimens showed polymicrbial growth. Overall, 69 pathogens were identified and characterized. Staphylococcus aureus (27.5%) was the predominant pathogen isolated. Methicillin resistance was noticed in seven strains of Stahylococcus aureus. Second predominant organism was Escherichia coli 14 (20.2%). Among fungi, Candida species was isolated and accounted for 6 (8.6%) as per **Table** -3.

<u>**Table – 3:**</u> Pathogens isolated from SSIs.

Isolate	Number (%)	
Staphylococcus aureus	19(27.5%)	
MSSA	12(63.1%)	
MRSA	7(36.8%)	
Escherichia coli	14(20.2%)	
Klebsiella species	11(15.9%)	
Pseudomonas aeruginosa	9(13%)	
Proteus species	7(10.1%)	
NFGNB	3(4.3%)	
Candida species	6(8.6%)	

(MSSA - Methicillin sensitive Staphylococcus aureus, MRSA - Methicillin resistant Sthaphylococcus aureus, NFGNB - Non fermenting gram negative bacilli)

All strains of Methicillin resistant staphylococcus aureus were susceptibility linezolide (100%) followed by co trimaxazole (71%) and cefeperazone/ sulbactum (71%). Methicillin sensitive staphylococcus aureus showed good susceptibility towards all antibiotics except cefotaxime (50%) and ofloxacin (50%). Seven strains of Klebsiella species were identified as extended spectrum of betalactamase (ESBL) producers. Among E.coli, six strains were found to be ESBL producers. Imipenem was the most active drug against ESBL strains followed by cefeperrazone/ sulbactum and piperacillin/ Tazobactum (**Table - 4**).

Antibiotics	MRSA	MSSA	E.coli	Klebsiella	Pseudomonas	Proteus	NFGNB
	(7)	(12)	(14)	(11)	(9)	(7)	(3)
Amoxyclav	3(42%)	12(100%)	12(85%)	9(81%)	8(88%)	7(100%)	3(100%)
Amikacin	3(42%)	11(91%)	10(71%)	8(72%)	9(100%)	7(100%)	1(33%)
Cefotaxime	1(14%)	6(50%)	6(42%)	5(45%)	7(77%)	3(42%)	0(0%)
Cotrimaxazole	5(71%)	11(91%)	7(50%)	5(45%)	3(33%)	5(71%)	0(0%)
Pipracillin/	4(57%)	12(100%)	9(64%)	11(100%)	9(100%)	7(100%)	1(33%)
Tazobactum							
Cefeperazone/	5(71%)	12(100%)	12(85%)	11(100%)	9(100%)	7(100%)	3(100%)
sulbactum							
Oflaxacin	4(57%)	6(50%)	6(42%)	7(63%)	2(22%)	4(57%)	1(33%)
Imipenem	NT	NT	13(92%)	9(81%)	9(100%)	7(100%)	2(66%)
Linezolide	7(100%)	12(100%)	NT	NT	NT	NT	NT

Table – 4: Antibiotic susceptibility pattern of bacterial isolates.

NT-Not tested

Discussion

In our study, surgical site infection rate was fond to 16%. This is similar to the study conducted by Brown, et al. (16.7%) [15] in republic of Georgia. In our study, Surgical site infection rate is very low compared to the studies conducted by Ganguly, et al. (38.8%) and Subramanian, et al. (24.8%) [7, 8]. But few studies from various parts of India showed low incidence of surgical site infection. Recent study by Chada, et al. showed very low incidence of surgical site infection rate (3.83%) [16]. Murthy R, et al. also reported 4.2% as incidence of SSI in their study [17].

Emergency surgeries were more likely to be associated with higher incidence of SSI in various studies done worldwide. The incidence of SSIs in our study was more in emergency surgeries 22/97 (22%) as compared to routine/ elective surgeries 33/245 (13%). Mahesh CB, et al. [18] also observed a similar SSI rate of 21.05% in emergency surgeries as compared to 7.61% of cases in elective surgeries. This can be due to the reasons like emergency surgeries which lack regular pre-operative preparation and involve mostly abdominal and intestinal surgeries which are contaminated surgeries.

In the present study, SSIs predominantly observed in age group above 65 years (46.42%) followed by 56-65 years (25%). This is similar to the study conducted by Astagneau, et al. [19]. In our study, when operation categorized by traditional wound classification, infections occurred in 7.9% of the clean wounds, 13.3% of the clean-contaminated wounds, 43.7% of the contaminated wounds and in 75.2% of the dirty or the infected wounds. These findings are similar to the study conducted by of Rosentha, et

al. in his study [20]. A study which was conducted at the Mayo Hospital, Lahore, reported an infection rate of 5.05% among the clean and a rate of 8.39% amongst the clean-contaminated cases [21].

Certain underlying conditions like anemia, diabetes, and smoking may alter or decrease the immune status thus significantly increasing the risk of SSI. They play the role in increasing the pre operative stay of the patient which steeply increases the risk of SSI in such patients. In our study, among diabetic patients SSI was found to be 23.9% and in non-diabetic patients 13.2%.

In the present study, Staphylococcus aureus was the predominant bacteria isolated and accounted for 27.5%. Staphylococcus aureus, is a major pathogen and a predominant cause of SSIs worldwide with a prevalence rate ranging from 4.6% to 54.4% [22]. Infection with S. aureus is most likely associated with endogenous source as it is a member of the skin and nasal flora and also with contamination from environment, surgical instruments or from hands of health care workers [23]. In the present study MRSA was accounted for 36.8%. This is low as compared to other studies conducted by Eagye, et al. (45%) and Kaye, et al. (58.2%) [24, 25]. All strains were found to be susceptible to linezolide (100%) followed by cotrimaxazole (71%).

Among Gram negative bacteria, E.coli (20.2%) and Klebsiella species (15.9%) were found to common pathogens followed by Pseudomonas aeruginosa (13.0%) Few studies have reported P.aeruginosa as the most frequent isolate in SSI [26]. All Gram negative bacilli showed good susceptibility to Imipenem, cefeperazone/ sulbactum and piperacillin/ tazobactum including ESBL producers. Candida species (spp) have emerged as the seventh most common health care - associated pathogen in the critically ill with an associated mortality rate of 19- 50% [27]. In the present study, 6(8.6%) of Candida species isolated.

In this study limitations are, only few factors which are associated with SSIs were taken into the study, no anerobic culture was done and only antibiotic susceptibility testing was performed. No antifungal susceptibility was done with Candida species isolated from SSIs.

Conclusion

In this study, surgical site infection rate was found to 16%. Association of various risk factors showed impact on developing SSIs. The rate of infection in diabetics and those who underwent emergency operations was significantly higher than others. Staphylococcus aureus (27%) was the most frequently isolated bacteria from SSIs. All MRSA strains were found to be susceptible to linezolide. Among Gram negative bacteria, E.coli (20%) was the predominant bacteria and showed good susceptibility to imipenem, cefeperazone/ sulbactum and piperacillin/ tazobactum including ESBL producers.

References

- Awad SS, Palacio CH, SubramanianA., Byers PA, Abraham P, Lewis D, et al. Implementation of a methicillinresistant Staphylococcus aureus (MRSA) prevention bundle results in decreased MRSA surgical site infections. Am J Surg., 2009; 198: 607-610.
- Horan TC, Gaynes RP, Martone WJ, Jarvis WR, Emori TG. CDC definitions of nosocomial surgical site infections, 1992: a modification of CDC definitions of surgical wound infections. Am J Infect Control., 1992; 20: 271-74.
- Kirkland KB, Briggs JP, Trivette SL, et al. The impact of surgical-site infections in the 1990s: attributable mortality, excess length of hospitalization, and extra costs. Infect Control Hosp Epidemiol., Nov 1999; 20(11): 725-30.
- Altemeier WA. Perspectives in surgical infections. Surg Clin North Am., 1980; 60: 5–13.

- Cruse Peter J, Foord R. The epidemiology of wound infection. Surg Clin North Am., 1980; 60(1): 27–40.
- Haley RW, Hooton TM, Culver DH, Stanley RC, Emori TG. Nosocomial infections in US hospitals, 1975–1976. Am J Med., 1980; 70: 947–958.
- Ganguly PS, Khan Y, Malik A. Nosocomial Infections and hospital procedures. Indian Journal of Community Medicine, 2000; 25(1): 39.
- Subramanian KA, Prakash A, Shriniwas, Bhujwala RA. Post-operative wound infection. Ind J Surg., 1973; 57–64.
- Mangram AJ, Horan TC, Pearson ML, Silver LC, Jarvis WR. Guidelines for prevention of surgical site infection. Infect Control Hosp Epidemiol., 1999; 20: 250-278.
- Howard RJ, Lee JT Jr. Surgical wound infections: epidemiology, surveillance, and clinical management. In Surgical Infectious Diseases, 3rd edition, Edited by: Howard RJ, Simmons RL. East Norwalk, CT: Allyn & Bacon; 1995, p. 401-412.
- 11. Mundhada AS, Tenpe S. A study of organisms causing surgical site infections and their antimicrobial in tertiary susceptibility а care Government Hospital. Indian J Pathol Microbiol., 2015; 58: 195-200.
- Bhatt C.P., Baidya R., Karki P., Shah R.K., Miya R., Mahashate P., Mishra K.K. Multi Drug Resistance Bacterial Isolates of Surgical Site Infection. Open Journal of Medical Microbiology, 2014; 4: 203-209.
- Oslon M, Lee JT Jr. Continuous, 10-year wound infection surveillance. Arch Surg., 1990; 125: 794–803.
- Wayne PA. Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing.21st information supplement M100- S21., USA: Clinical and laboratory standards institute, 2011.

- 15. Brown S, Kurtsikashvili G, Alonso-Echanove J, Ghadua M, Ahmeteli L, Bochoidze T., et al. Prevalence and predictors of surgical site infection in Tbilisi, Republic of Georgia. J Hosp Infect., 2007; 66: 160-166.
- Chada CKR, Kandati J, Ponugoti M. A prospective study of surgical site infections in a tertiary care hospital. Int Surg J., 2017; 4: 1945-52.
- Murthy R, Sengupta S, Maya N, Shivananda PG. Incidence of postoperative wound infection and their antibiogram in a teaching and referral hospital. Indian J Med Sci., 1998; 52: 553-5.
- Mahesh CB, Shivakumar S, Suresh BS, et al. A Prospective study of surgical site infections in a teaching hospital. J Clinical Diagnostic Research, 2010 Oct; 4(5): 3114-9.
- Astagneau P, Heriteau FI, Daniel F, Parniex P, Venier AG, Malvaud S, et al. Coignard for the ISO-RAISIN Steering Group. Reducing surgical site infection incidence through a network: results from the French ISO-RAISIN surveillance system. J Hosp Infect., 2009; 72: 127-134.
- Rosenthal R, Weber WP, Marcel Z, Misteli H, Reck S, Oertli D, et al. Impact of surgical training on incidence of surgical site infection. World J Surg., 2009; 33: 1165-73.
- 21. Akhtar S, Gondal KM, Ahmed M, Mohammad Y, Goraya AR, Karim K, Chaudhry AM. Surgical Wound Site Infections-Our experience. Ann King Edward Med Coll., 2001 Sept; 7(3): 211-12.
- Chakarborty SP, Mahapatra SK, Bal M, Roy S. Isolation and identification of vancomycin resistant Staphylococcus aureus from postoperative pus sample. Al Ameen J Med Sci., 2011; 4(2): 152-68.
- 23. Isibor OJ, Oseni A, Eyaufe A. Incidence of aerobic bacteria and Candida albicans

in postoperative wound infections. Afr J microbial Res., 2008; 2: 288-91.

- Eagye KJ, Kim A, Laohavaleeson S, Kuti JL, Nicolau DP. Surgical site infections: does inadequate antibiotic therapy affect patient outcomes? SurgInfect (Larchmt), 2009; 10(4): 323-31.
- 25. Kaye KS, Anderson DJ, Sloane R, Chen LF, Choi Y, Link K, et al. The effect of surgical site infection on older operative

patients. J Am Geriatr Soc., 2009; 57(1): 46-54.

- Masaadeh HA, Jaran AS. Incident of Pseudomonas aeruginosa in postoperative wound infection. Am J Infect Dis., 2009; 5: 1–6.
- 27. Bustamante CI. Treatment of Candida infection: a view from the trenches! Curr Opin Infect Dis., 2005; 18: 490-5.