

Original Research Article

Characterisation of Susceptibility of Candida Spp. to Three Essential Oils – A Study done in FIMS, Kadapa

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Abstract

In the present investigation, anti-Candida activity of three essential oils that is Betel leaf (*Piper betel*), Black cumin (*Nigella sativa*) and Curry leaf (*Murraya koenigii*) were screened against three human pathogenic species of Candida namely *Candida albicans*, *Candida glabrata* and *Candida tropicalis*. The minimum inhibitory concentration (MIC) values of the oils ranged between 14.80 and 236 µl/ml while studied through the dilution method. The oils retained their anti-Candida activities even after heat treatment (at 45⁰C, 60⁰C, 100⁰C for 1 hour) and also on autoclaving. Black cumin leaf oil showed better anti-Candida activity against *Candida albicans*, resulting in an irreversible damage to the cells. The anti-Candida activity of these essential oils could be attributable to the membrane inhibition mechanism. The activity of the cells is reported to be microbicidal.

Key words

Anti-Candidal activity, Essential oils, Minimum inhibitory concentration.

Introduction

For the past ten years fungal infections are increasing and a rise in resistance to fungicides

for most of the species are seen in routine medical practice. Of all the hospital acquired blood stream infections Candida species

infections are fourth leading cause. Leading to mortality of up to 40% for systemic and disseminated infections. Clinical manifestations include oropharyngeal infections and infections among persons with Human Immunodeficiency Viruses (HIV) or full blown disease of Acquired Immunodeficiency Syndrome (AIDS) patients and also candidaemia, vulvovaginal infections affecting women of all age groups [1-5].

About 50% of both superficial and systemic mycoses and almost all the mucosal candidiasis is caused by *Candida albicans*. However serious oropharyngeal candidiasis and occasional esophageal candidiasis were caused by non albican Candida species such as *Candida glabrata*, *Candida tropicalis*, *Candida parapsilosis* and *Candida krusei*.

There is increased interest generated among the academicians and researchers towards herbal medications because of development of drug resistance towards the commonly used antibiotics and chemotherapeutic agents. For centuries, the therapeutic properties of various medicinal plants have been used to treat human diseases. It has been estimated that between 50% to 80% of the populations of developing countries use traditional and botanical medicines almost exclusively and consider them to be a normal part of primary healthcare [6-11]. Essential oils derived from plants are well known in traditional medicine and proved to have insecticidal, bactericidal, fungicidal and nematocidal effects. Antibacterial activity of essential oils are well cited in literature. However, the antifungal activity of essential oils specifically against Candida species is less studied. Because of that this study has been done. So, a study in Fathima Institute of Medical Sciences, Kadapa was done in September 2013.

Materials and methods

- Betel leaf (*Piper betel*)
- Black cumin (*Nigella sativa*)
- Curry leaf (*Murraya koenigii*)

- Hydro-steam distilled essential oils derived from Southern Spice Pvt. Ltd. Madurai and used for this study.
- Fungal Pathogens: -
- Human pathogenic species of Candida such as –
- *Candida albicans*
- *Candida glabrata* and
- *Candida tropicalis*

Above species were maintained on Sabouraud's Dextrose Agar slants in the laboratory.

Minimum Inhibitory Concentration (MIC) – Determination

Minimum Inhibitory Concentration (MIC) of the oils were determined by double dilution method [14] in Sabouraud's Dextrose Broth supplemented with Tween 20 (0.75%) to facilitate miscibility of oils.

Minimum Killing time of Oils – Determination

To determine the Minimum Killing Time of the oils against the test pathogens, 1 ml of Sabouraud's Dextrose Broth with Tween – 20 (0.75%), at the MIC level of the oils was prepared and inoculated with 0.1 ml of freshly grown test organisms and incubated at 5⁰C, room temperature (25±2⁰C) and 37⁰C respectively. One loopful of the sample from the above test tubes were subcultured onto SDA plates at 0, 5, 10, 15, 30, 45, 60, 120, 180, 240, 300, 360, 420, 480 minutes intervals and incubated overnight. Two sets of tubes were incubated for each test organism at a specific temperature from which subculturing was carried out alternatively to minimize the time lapse during subculture. The activity was observed after overnight incubation of the plates at 25⁰C±2⁰C. Absence of growth on the streak line was considered to be the time taken by the oil to kill the organism.

Effect of pressure and temperature on anti-Candida activity of the oils – Determination

The effect of pressure and temperature was studied by autoclaving at 121⁰C for 20 minutes and heating the oils at different temperatures that

is 45⁰C, 60⁰C and the activity was studied at MIC levels of the oils.

Results

Minimum Inhibitory Concentration

Minimum Inhibitory Concentration (MIC) of oils ranged from 14.80 to 236 µl / ml (**Table – 1**).

Lowest MIC value of 14.8 µl/ml was reported in case of *Nigella sativa* oil against *Candida albicans* and *Candida tropicalis*. *Murraya koenigii* showed MIC value 128 µl / ml against *Candida albicans* and *Candida glabrata* whereas, the same oil showed higher MIC values of 236 µl / ml against *Candida tropicalis*.

Table – 1: Minimum Inhibitory Concentration of the oils against test pathogens.

Pathogens	MIC (µl / ml) of oils		
	<i>Piper betel</i>	<i>Nigella sativa</i>	<i>Murraya koenigii</i>
<i>Candida albicans</i>	127.0	14.80	128.0
<i>Candida glabrata</i>	58.25	42.15	126.2
<i>Candida tropicalis</i>	28.70	14.80	236.0

MIC: Minimum Inhibitory Concentration

Fungicidal / Fungistatic nature of oils – Determination

By subculturing one loopful of the sample from the MIC dilution tubes onto Sabouraud's Dextrose Agar plates an attempt was made to study the fungicidal/ fungistatic nature of the oils. It was observed that all the oils showed the fungicidal (Candidacidal) nature, as no growth was observed on the Sabouraud's Dextrose Agar plates after the incubation period.

Minimum Killing Time

An attempt was made to study the time required to kill the pathogens at a particular temperature at MIC level of oils. A variation was observed in Minimum Killing Time of the oils when tested at three different temperatures (**Table – 2**). Immediate killing of *Piper betel* oil was observed against *Candida albicans* at all the three temperatures studied. However, similar effect of *Piper betel* oil was also observed against *Candida glabrata* at 37⁰C and *Candida albicans* at 5⁰C, respectively.

Effect of temperature and pressure on anti-candida activity of the oils

This experiment was designed to study the effect of temperature and pressure on the anti-Candida activity of the oils. Oils were heated in a boiling water bath (45, 60, 100⁰C for 1 hour and

autoclaved at 121⁰C and 15 lb pressure for 20 minutes) and studied for their anti-Candida activities by Disc Diffusion Method loading MIC values of the oils onto the discs. Surprisingly, a significant increase in the activity of the oils was reported due to heating in case of *Piper betel* and *Nigella sativa* that showed complete inhibition of the test pathogens (pathogens showing poor growth at the centre of the Petri dish and no growth around the discs), on SDA plates.

Discussion

Three different essential oils were tested for their antimicrobial properties against *Candida* species. During the investigation we observed lower MIC values of test essential oils against the *Candida* species which is indicative of their high degree of effectiveness against these pathogens. Observance of lower MIC value of essential oils against both bacteria and fungi is being reported in literature. In contrast to the findings observed in this investigation Rath, et al. (1999) reported a higher MIC values 62.5 and 500 µl / ml of turmeric leaf and rhizome essential oils respectively, against *Candida albicans*. While studying the microbicidal/ microbiostatic nature of these oils, the oils were reported to be microbicidal (*Candida*-cidal). Microbicidal nature of different essential oils is well documented in literature [12-20].

Table – 2: Minimum Killing Time of the Oils.

Pathogens	Minimum Killing Time of the Oils in minutes								
	<i>Piper betel</i>			<i>Nigella sativa</i>			<i>Murraya koenigii</i>		
	5 ⁰ C	RT	37 ⁰ C	5 ⁰ C	RT	37 ⁰ C	5 ⁰ C	RT	37 ⁰ C
<i>Candida albicans</i>	0	0	0	130	100	110	160	30	10
<i>Candida glabrata</i>	30	110	0	110	110	100	170	160	20
<i>Candida tropicalis</i>	320	160	110	280	50	50	110	40	20

During the determination of Minimum Killing Time of the oils it was observed that *Piper betel* killed *Candida albicans* immediately at all three test temperatures. Similar results were also observed for *Piper betel* against *Candida glabrata* at 37⁰C. Killing of the *Candida* species immediately by these oils indicates that the oils cause an irreversible damage to the structure of the test organisms when they come in contact with the oil mixture. Immediate killing of pathogens and irreversible damage to cellular structure by essential oils is well recorded in literature. Since the oils showed activity against *Candida* spp. at both lower temperatures 5⁰C and 37⁰C, it implies that the activities of these oils are energy independent [21-24].

Energy-independent bactericidal activities of lemongrass, palmarosa and eucalyptus oil against *Escherichia coli* and other bacteria is also recorded. Rath, et al., (2001) reported similar observations while studying the antifungal activity of fractionally distilled and neat turmeric leaf oil. They reported that neat oil killed *Candida albicans*, *Cryptococcus neoformans*, *Trichophyton rubrum* and *Microsporum gypsum* within 1 minute of treatment whereas, 1 hour and 2 hour fractionally distilled oil took a longer time to kill the same pathogens in comparison to neat oil. Our observations here can be agreed with Rath et al., (1999) reported immediate killing of *Candida albicans* and *Cryptococcus neoformans* when treated for a longer time (15 min) in comparison to turmeric leaf oil .

Even on heating the oils at 45, 60, 100⁰C for 1 hour and autoclaved at 121⁰C and 15 lb pressure for 20 minutes, anti-Candidal activity was not

lost. Similar properties of various essential oils have been reported by researchers while studying their antimicrobial activities including *Candida* species. On heat treatment, a significant increase in anti-*Candida* activity of *Nigella sativa* and *Piper betel* oils was recorded during the investigation. This could be attributable to the change in charge of the compounds present in these essential oils and increase in their mobility.

Further, it suggests that the anti-*Candida* components that are present in these oils are heat stable and withstand a temperature of 121⁰C and 15 lb pressure, indicating their thermostable and barostable nature. Gupta et al., (2004) reported the persistence of antimicrobial activity of carrot (*Daucus carota*) and celery (*Apium graveolens*) seed essential oils against both Gram positive and Gram negative pathogens after heat treatment (100⁰C) and autoclaving, indicating the presence of heat stable components in these essential oils as reported in our investigation. Similar results are also reported in literature while studying the antibacterial activity of lime (*Citrus limonum*) and juniper (*Juniperus communis*) oils against 32 strains of methicillin resistant *Staphylococcus aureus*. Das et al., (2009) reported the antibacterial activity of essential oils of three *Ocium* spp. and their cocktail mixture at high temperature and pressure concluding the presence of heat stable and barostable components in essential oils, as reported here in our studies.

Pathogen susceptibility to the oils that are tested may be due to inhibition of cell membrane synthesis, specifically by extracting the sterols from the membrane or inhibiting steroid

synthesis. Senhaji et al., (2007) observed the antibacterial activity of essential oil from *Cinnamum zeylanicum* against *Escherichia coli* 0157:H7 is through outer membrane disintegration and increasing the permeability of ATP through cytoplasmic membrane. Similarly, Rath et al., (2005) also reported the anti-Staphylococcal activity of juniper and lime essential oils against methicillin – resistant *Staphylococcus aureus* (MRSA) through inhibition of cell membrane synthesis. Further, it is to add that the essential oils are rich in terpenes. However, the mode of action of terpenic constituents (essential oils) on microorganisms is not fully understood. But, in view of their hydrophobicity, it is considered that they are involved in mechanism such as permeability of cytoplasmic membrane, coagulation of cell contents and disruption of the proton motive force. Therefore, the anti-Candida activity of these essential oils through membrane inhibition could be attributable to the hydrophobicity of essential oils that enables them to make partitions in the membrane, rendering permeability due to extraction of steroid molecules present on the membrane and leading to leakage of cell contents resulting in death of the cells [25-28].

Conclusion

With this study we came to a conclusion that anti-Candidal activity of these essential oils against human pathogens is suggestive of their use in pharmaceuticals and cosmetic industries for production of drugs and aroma products. However, further scientific research is essential to investigate the side effects of these oils before consideration of their use.

References

1. Garbino J, Adam A. Use of high dose liposomal amphotericin B: Efficacy and tolerance. *Acta Biomed.*, 2006; 77: 19-22.
2. Garber G. An overview of fungal infections. *Drugs*, 2001; 61: 1-12.
3. Pfaller MA, Diekema DJ. Epidemiology of invasive candidiasis: A persistent public health problem. *Clin Microbiol Rev.*, 2007; 20: 133-63.
4. White TC, Marr KA, Bowden RA. Clinical, cellular and molecular factors that contribute to antifungal drug resistance. *Clin Microbiol Rev.*, 1998; 11: 383-402.
5. Nace HL, Horn D, Neofytos D. Epidemiology and outcome of multiple-species candidemia at a tertiary care center between 2004 and 2007. *Diagn Microbiol Infectious Dis.*, 2009; 64: 289-94.
6. WHO. 2002. Traditional Medicine Growing Needs and Potential – WHO Policy Perspectives on Medicines, No. 002, May 2002, World Health Organisation, Geneva, Switzerland; 2002, p. 1-6.
7. Wingard JR. Importance of Candida species other than *C. albicans* as pathogens in oncology patients. *Clin Infect Dis.*, 1995; 20: 115-25.
8. Vazquez JA, Arganoza MT, Boikov D, Akins RA, Vaishampayan JK. In vitro susceptibilities of Candida and Aspergillus species to Melaleuca alternifolia (tea tree) oil. *Rev Microbiol.*, 2000; 17: 60-3.
9. Rath CC. Recent progresses in medicinal plants: Pharmacology and Therapeutic Values III. In: Singh VK, Govil JN, editors. *Essential oils: Their Role in Antimicrobial Activities and Aromatherapy an Overview*. 1st edition. Vol. 21. LLC: Studium Press; 2008, p. 63-87.
10. Patil SD, Kamble VA. Antibacterial activity of some essential oils against food borne pathogen and food spoilage bacteria. *Int J Pharm Biosci.*, 2011; 2: 143-50.
11. Werdin Gonzalez JO, Gutierrez MM, Murray AP, Ferrero AA. Composition and biological activity of essential oils from Labiatae against *Nezera viridula*

- (Hemiptera: Pentatomidae) soyabean pest. Pest Manag Sci., 2011; 67: 948-55.
12. Zabaka M, Pavela R, Slezakova L. Antifungal effect of *Pinmentia dioica* essential oil against dangerous pathogenic and toxigenic fungi. Ind Crops prod., 2009; 30: 250-3.
 13. Kamble VA, Patil SD. Spice-derived essential oils: Effective antifungal and possible therapeutic agents. J Herbs Spices Med Plants, 2008; 14: 129-43.
 14. Das M, Rath CC, Mohapatra UB. Bacteriology of a most popular street food (panipuri) and inhibitory effect of essential oils on bacterial growth. J Food Sci Technology, 2012; 49: 564-71.
 15. Leela NK, Ramana KV. Nematicidal activity of the essential oils of all spice (*Pimenta dioica* L. Merr.). J. Plant Biol., 2000; 27: 75-6.
 16. Burt S. essential oils: Their antibacterial properties and potential applications in food-a review. Int J Food Microbiol., 2004; 94: 223-53.
 17. Gochhait S, Rath CC, Mohapatra UB. In: Gupta VK, Singh GD, Kaul A, editors. Medicinal Plants: Phytochemistry, Pharmacology and Therapeutics. Antibacterial Properties of Some Essential Oils Against Selective Human Pathogenic Bacteria. 1st edition. Vol. 2., India: Daya Publishing House; 2012, p.379-88.
 18. Behera R, Rath CC. Evaluation of antimicrobial activity of turmeric (*Curcuma longa* L.) leaf essential oils of three different states of India against *Shigella* spp. J Biol Active Plant Prod Nat., 2011; 1: 125-31.
 19. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disc method. Am J Clin pathol., 1996; 45: 493-6.
 20. Rath CC, Das SK, Mishra RK, Charuyulu JK. In vitro evaluation of antimycotic activity of turmeric (*Curcuma longa* L) essential oil against *Candida albicans* and *Cryptococcus neoformans*. Ind Perf., 1999a; 43: 172-8.
 21. Gupta R, Rath CC, Dash S, Mishra RK. In vitro antibacterial potential assessment of Carrot (*Daucus carota*) and Celerey (*Apium graveolens*) seed essential oils against twenty one bacteria. J Essent Oil Bear PI, 2004; 7: 79-86.
 22. Pattnaik S, Suvramanyam VR, Rath CC. Effect of essential oils on the viability and morphology of *Escherichia coli* (SP-11). Microbios., 1995; 84: 195-9.
 23. Rath CC, Mishra S, Dash SK, Mishra RK. Antistaphylococcal activity of lime and juniper essential oils against MRSA. Ind Drugs, 2005; 42: 797-801.
 24. Aghel N, Mahmoudabadi AZ, Darvishi L. Volatile constituents and anti-candida activity of the aerial parts essential oil of *Dittrichia graveolens* (L.) Greuter grown in Iran. Afr J Pharmacol., 2011; 5: 772-5.
 25. Rath CC, Das SK, Mishra RK. Antibacterial efficacy of six Indian essential oils individually and in combination. J Essent Oil Bear P1, 2002; 5: 99-107.
 26. Das I, Tayung K, Rath CC, Mohapatra UB. Antibacterial assessment of essential oils of three *Ocimum* spp. against food borne pathogens. Plant Sci Res., 2009; 31: 60-5.
 27. Rath CC, Das SK, Mishra RK, Ramachandraiah GS. A Comparative analysis of in vitro antifungal activity of pure and fractionally-distilled turmeric (*Curcuma longa*) leaf essential oil. Ind Drugs, 2001; 39: 18-22.
 28. Kumar A, Thakur S, Thakur VC, Kumar A, Patil S, Vohra MP. Antifungal activity of some natural essential oils against *Candida* species isolated from blood stream infections. J Krishna Inst Med Sci Univ., 2012; 1: 61-6.